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RARE 2-SUBSTITUTED PURINE NUCLEOSIDES

ANNUAL/FINAL REPORT

May 1989

Vasu Nair

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The University of Iowa Iowa City, Iowa 52242

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1. Project Title: "Rare 2-Substituted Purine Nucleosides"

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4. Reporting Period: October 18, 1985 to April 17, 1989

5. Description of Work Accomplished:

The overall goals of the contract were to develop procedures and synthesize, purify, and submit with complete physical data sixteen rare purine nucleosides for antiviral studies. These goals were accomplished. A total of twenty-two novel compounds were submitted to the Department of Antiviral Studies, USAMRIID for biological evaluation. A description of the synthetic work accomplished during the entire period of the award in chronological order, the compounds submitted, the biological screening data, publications, personnel supported, and an executive summary are given in the pages that follow.

In the first year of this contract, our goals were to develop rational procedures for the synthesis of the target molecules discussed in the proposal. Although difficulties were encountered in approaches studied in the first six months of the contract, these problems were entirely overcome and an excellent and novel methodology for the key step in the synthesis of many of the target compounds was discovered.

The starting point of our work during the first year involved the preparation of multigram quantities of some basic precursors and investigation of feasible approaches to the synthesis of 2-acetonylnebularine 1 and 2-acetonylinosine 2. Target molecules 1 and 2 (in their protected form) were considered to be direct precursors for the synthesis of 3 and 4, respectively.

The synthesis of compound 1 commenced with guanosine (5) which was converted almost quantitatively (93%) in the first step to 2',3', 5'-tri-0-acetylguanosine (6) by treatment with acetic anhydride, dimethylaminopyridine, and triethylamine in acetonitrile as solvent. The protected nucleoside 6, when treated with phosphorus oxychloride and N,N-diethylamiline, was converted

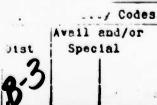
to the 2-amino-6-chloropurine nucleoside 7 in about 89% yield. Photolysis of 7 in dry, nitrogen-purged tetrahydrofuran (THF) containing 10% triethylamine produced the 2-amino nucleoside 8 in about 80% yield. This photoinduced reductive dehalogenation has not been reported previously in purine nucleoside chemistry and represents an excellent procedure for the synthesis of 2-aminopurines. Nucleoside 8 was designed as a key precursor for the synthesis of 1 and 3.

Several approaches were examined for the synthesis of f 1 from f 8.first involved conversion of 8 to its 2-iodinated derivative with subsequent photoinduced Spul reaction of this iodo compound with the potassium enolate of acetone (Scheme 1). Nucleoside 9 (i.e. the silylated derivative of 8) was converted to the new 2-iodo-9-(2,3,5-tri-0-t-butyldimethylsilyl-D-ribofuranosyl)purine 10 in 67% yield by a deamination-halogenation reaction using n-pentyl nitrite, diiodomethane, and trimethylsilyl iodide in hexane. However, when 10 was photolyzed in the presence of the potassium enolate of acetone in THF at -48°C for 20 minutes, the expected SRN1 product was not isolated. Careful analysis of the high-field NMR, FTIR, UV, and mass spectral data suggested that nucleophilic attack with subsequent ring opening had occurred at the 6-position to produce 11. Although this was totally unexpected, failure of the S_{RN}l reaction may be attributed to a marked change in the reduction potential of 10 compared to the corresponding 6-iodo compound. The latter undergoes the SRN1 reaction in very good yields. When the 6-position was blocked, as in the case of 2-iodo-6-methoxypurine nucleoside (a precursor for target molecule 2), the reaction still failed.

Scheme 1

An alternative approach to compound 1 also involved the use of the 2-amino nucleoside 8 as a precursor. The methodology involved conversion of 8 to its 2-thioacetonyl derivative and subsequent application of the Eschenmoser sulfide contraction on this thio derivative (Scheme 2). Nucleoside 8 can be converted to its 2-thioacetonyl derivative 12 in good yields by heating with n-pentyl nitrite and diacetonyl disulfide in acetonitrile. The Eschenmoser sulfide contraction reaction on 12 to give 14 via the intermediacy of 13 did not proceed as planned under a variety of conditions. Modifications in the experimental procedure included changing the solvent, the base, and the phosphine used.





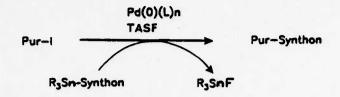
Scheme 2

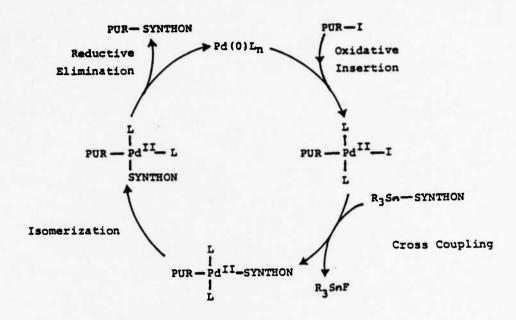
A methodology involving radical addition of thioacetone at the 2-position of an appropriate precursor followed by thermal sulfide contraction and subsequent modification at the 6-position appeared also to be a promising and direct approach to the synthesis of 2 (Scheme 3). However, the sulfide contraction was also unsuccessful in this case. Application of the Meerwein reaction and radical coupling reactions did not provide suitable routes to 1 and 2.

$$7 \longrightarrow \bigvee_{\substack{S \\ CH_1 \\ C = 0 \\ CH_1}} \bigcap_{\substack{R \\ C \\ CH_2 \\ CH_3}} \bigcap_{\substack{R \\ C \\ CH_3 \\ CH_3}} \bigcap_{\substack{R \\ C \\ CH_3 \\ CH_3}} \bigcap_{\substack{R \\ CH_3 \\ CH_3 \\ CH_3}} \bigcap_{\substack{R \\ CH_3 \\ CH_3 \\ CH_3}} \bigcap_{\substack{R \\ CH_3 \\ CH_3$$

Scheme 3

Palladium is known to be able to insert into the carbon-iodine bond of an iodoaromatic system and this intermediate can subsequently undergo cross-coupling reactions with alkenes under suitable conditions. We have been very successful in developing this methodological approach for the synthesis of a number of the target nucleosides for this contract. The conceptual approach and a mechanistic explanation of the reaction is shown in Scheme 4. This conversion involves insertion of palladium into the carbon-iodine bond of the iodopurine followed by coupling of the derived Pd(II) complex with the tin enolate of acetone, trans-cis isomerization, and reductive elimination to give the product with regeneration of the Pd(0). Only catalytic amounts of palladium are required for this reaction. It is the first example of the use of an Sn reagent in palladium-catalyzed coupling involving nucleosides.





PUR-I - Iodopurine Nucleoside System

Scheme 4

The methodology is illustrated with the synthesis of 2 (Scheme 5). The synthesis commenced with guanosine (5) which was converted to the 2-amino-6-chloropurine nucleoside 7 in excellent yield as previously described in this report. Reaction of compound 7 with n-pentyl nitrite and diiodomethane in refluxing acetonitrile gave the 6-chloro-2-iodopurine nucleoside 18 in 71% yield. Replacement of the chlorine group at the 6-position with methoxide is accompanied by the desired deprotection of the acetate groups to give 19 in 76% yield. Subsequent protection of the carbohydrate moiety with t-butyldimethylsilyl chloride and imidazole in dimethylformamide gave 20 in 96% yield. The key step in the synthesis of 1 was the conversion of 20 to 21 in 70% yield by the palladium catalyzed coupling reaction discussed above. The acetonylated nucleoside 21 was converted to 2 in two steps by reaction first with trimethylsilyl iodide (64% yield of 22) and subsequently with tetrabutyl-ammonium fluoride (93% yield). The overall yield of 2 starting from guanosine was an excellent 17.9%.

Compound 2 was purified to a high degree by reversed-phase high performance liquid chromatography (3 passes) on Amberlite XAD-4 resin using water-ethanol as the eluting solvent. The product may be crystallized from a water-isopropanol mixture. Complete characterization was performed by mass spectrometry, UV, FTIR, and high-field H and 13C NMR spectroscopy. Critical spectral data were presented with the sample submitted for antiviral testing.

Scheme 5

The starting material for the synthesis of compound 4 was the acetonylated nucleoside 21 (Scheme 5). It was smoothly reduced with sodium borohydride in tetrahydrofuran to give the diastereoisomeric products 23 in 75% purified yield. Compound 23 was deprotected to 4 in two steps by reaction first with trimethylsilyl iodide and subsequently with tetrabutylammonium fluoride. The overall yield of 4 starting from guanosine was an excellent 13%. Compound 4 was purified by multiple reversed phase HPLC on Amberlite XAD-4 with 5% ethanol-water as the eluting solvent. Crystallization may be

carried out from isopropanol. Complete characterization was carried out by mass spectrometry, UV, FTIR, and high-field H and 13C NMR spectroscopy. Complete spectral data were provided with WRAMC FORM 108.

Using the aforementioned methodology, the preparations of 2-acetony1-9-(\$\beta\$-D-ribofuranosy1)purine 1 and 2-(2-hydroxypropy1)-9-(\$\beta\$-D-ribofuranosy1)purine 3 were achieved. The approaches used for these compounds are outlined in Scheme 6. The precursor material for the syntheses was 10, prepared from 7 as described previously (Scheme 1). The palladium-catalyzed reaction of 10 with the tributyltin enolate of acetone gave 24 in good yields. Compound 24 can be easily deprotected to 1 with tetrabutylammonium fluoride. It can be reduced with sodium borohydride to 25 which can be deprotected to 3. Both compounds 1 and 3 were fully characterized and submitted for antiviral evaluation.

Scheme 6

In the second year of this contract, our goals were to utilize the procedures previously developed to synthesize a large number of target molecules. A total of nine rare C-2 functionalized nucleosides (target compounds) were submitted to the Department of Antiviral Studies for biological evaluation between October 18, 1986 and October 17, 1987.

The starting point of our work during the second year of the contract was the synthesis of the 2-vinyl compounds 26 and 27. The rationale for the choice of these compounds as the starting point was that, in addition to being target molecules, they would also be key precursors for the synthesis of a variety of rare functionalized alkylated purine nucleosides.

The synthesis of 2-vinylinosine (26) commenced with guanosine (5) which was converted in several steps, as described previously in the report, to 2-iodo-6-methoxypurine nucleoside (19) (Scheme 7). Reaction of 19 with tri-n-

butylvinylstannane in the presence of palladium chloride gave 28 in > 90% yield. It should be noted that the palladium-catalyzed cross-coupling reaction was carried out on nucleoside 19 where the carbohydrate moiety was totally unprotected. Deprotection of 28 with trimethylsilyl iodide in acetonitrile resulted in cleavage of the methyl group to give the target molecule 26 in about 50 % yield after appropriate work up and purification. Our procedure for masking the hypoxanthine base in this way will find wide application in purine nucleoside chemistry. Compound 26 was purified by high performance liquid chromatography (three passes) on Amberlite XAD-4 resin with ethanol-water as the eluting solvent. Complete characterization was carried out by UV, FTIR, high-field ¹H and ¹³C NMR spectroscopy, and elemental analysis.

Compound 27 was synthesized using the sequence of reactions shown in Scheme 8. The starting material for the synthesis was the 6-chloropurine

Scheme 8

nucleoside 7 which was converted in several steps to the 2-iodopurine nucleoside 10 as shown in Scheme 8 and as described previously in this report. Reaction of 10 with tri-n-butylvinylstannane under palladium catalysis gave the 2-vinylpurine nucleoside 29 which was deprotected with tetrabutylammonium fluoride to the target molecule 27. The overall yield starting from guanosine was 12%. The crude product was purified by flash chromatography followed by HPLC, fully characterized by spectral data, and submitted for antiviral evaluation.

Hydroxylation of compound 28 with osmium tetroxide gave the 1,2-dihydroxyethyl compound 30 in 55 % yield. Deprotection of 30 with trimethylsilyl iodide in acetonitrile gave 31 in 50 % yield (Scheme 9). This target molecule (obtained as a diastereoisomeric mixture) was purified by HPLC on Amberlite XAD-4 resin. It was fully characterized by spectral methods and elemental analysis. In a similar procedure, the vinylnebularine 27 was converted to the diastereoisomeric diols 32.

Scheme 9

The 2-vinyl purine nucleosides described could be elaborated further by selective oxidation at the beta-position to ful ish other target molecules. The methodology for this conversion is outlined in Scheme 10. 9-Borabicylco [3.3.1]nonane (9-BBN) is expected to add regiospecifically to the vinyl group of 33 to give the borane 34 with the boron bearing moiety at the terminal position. Oxidation of 34 with alkaline hydrogen peroxide would furnish the desired alcohol 35. It was planned that target molecules 35 would be precursors for the corresponding aldehydes 36 via controlled oxidation. With respect to the hydroboration reaction, it should be mentioned that such reactions have rarely been used to elaborate structures in nucleoside chemistry.

Pur-CH=CH₂ +
$$\frac{H}{33}$$

Pur-CH₂-CH₂- $\frac{G}{34}$
 $\frac{1}{34}$
 $\frac{1}{4}$

Pur-CH₂-CH₂OH

 $\frac{35}{36}$

Pur-CH₂-CC

 $\frac{1}{4}$
 $\frac{1}{36}$

Scheme 10

Thus, the rare functionalized inosine analogue, 2-(2-hydroxyethyl)inosine 40, was synthesized from the 2-vinyl compound 37. This compound was first protected by silylation to give 38, which was then treated with 9-borabicyclo [3.3.1]nonane (9-BBN). Oxidative work-up of the resulting organoborane gave the alcohol 39 in 52% yield. High-field H NMR spectral data confirmed that the regiospecificity of reaction as well as the structure of the isomer isolated. Deprotection of 39 with trimethylsilyl iodide in acetonitrile

followed by treatment with fluoride ions gave the target alcohol 40 (Scheme 11). Purification and characterization were carried out as described for other target compounds.

Scheme 11

In the last part of the second year of support, four compounds were submitted for antiviral evaluation. Two of the compounds, 41 and 42, were prepared by deprotection of key halogenated intermediates used in the syntheses previously described in this report. The other two compounds were target ketones 44 and 46 in which special emphasis was placed because of the potent antiviral activity of another ketone, 2-acetonylinosine, previously synthesized by us in this program.

The immediate precursor for the synthesis of the 2-ketoinosine 44 was the silylated 2-iodo compound 20. When compound 20 was heated under reflux in toluene with palladium acetate, tri-o-tolylphosphine, tri-n-butyltin methoxide, and 2-pentene-3-acetate, very good yields of the keto compound 43 was obtained. The latter was deprotected to the target molecule 44 in two steps, first by reaction with trimethylsilyl iodide and then with tetraethylammonium fluoride (Scheme 12). Target compound 44 was purified by reversed-phase HPLC. The overall yield of 44 starting from guanosine was 10.8%. It was fully characterized by spectral methods and by high-resolution fast atom bombardment mass spectrometry (FAB HRMS).

Synthesis of the ketonebularine 46 was achieved using the silylated 2-iodopurine nucleoside 10 as the immediate precursor. The palladium-catalyzed cross-coupling reaction of 10 to give 45 was carried out as described above for the conversion of 20 to 43. Excellent yields of product were obtained in this conversion. Deprotection of 45 (tetraethylammonium fluoride) followed by purification of the resulting material by HPLC, gave target molecule 46 (13.7% overall yield from guanosine).

Scheme 13

Epoxy substituted purine nucleosides are very rare compounds and only one example of a purine system with an epoxy group at the 6-position is known (Nair and Chamberlain, J. Am. Chem. Soc. 1985, 107, 2183). The approach to the 2-epoxy compounds of the inosine and nebularine series was through the corresponding vinyl compound precursors (38 and 29) whose synthesis have been described previously in this report. Although epoxidation of these vinyl compounds appeared to have proceeded as expected to give the epoxides 47 and 48 (Scheme 14), isolation of the epoxide products was extremely difficult because of their inherent instability. Several different procedures for isolation and deprotection were attempted, but all were unsuccessful.

Scheme 14

During the second year of the contract, we were also involved in the development of approaches to the synthesis of analogues of nebularine and inosine that contain aldehyde functionalized carbon-carbon bonding at the 2position. The initial approach for the nebularine series was to synthesize the 2-(2-hydroxyethyl)purine system and selectively oxidize this primary hydroxyl group to the aldehyde. Although the synthetic procedure for the preparation of the 2-hydroxyethyl derivative of inosine had previously been developed by us, application of this to the nebularine series (i.e. hydroboration followed by oxidative work-up) resulted in the formation of the 2-ethyl compound through reduction of the intermediate organoborane. alternative procedure involved direct introduction of a masked aldehyde moiety at the C-2 position. This was achieved through the use of ethyl vinyltributyltin ether. This organostannane was prepared by the radical coupling of tributyltin hydride with ethyl ethynyl ether. Palladium-catalyzed coupling of the organostannane with protected 2-iodopurine nucleoside 10 gave the (E)- and (Z)- mixture of the expected product 49 in about 70% yield (Scheme 15). However, although removal of the silyl protecting groups from 49 could be achieved to give 50, attempted unmasking of vinyl ether group gave an intractable mixture under a variety of conditions. The same problem was encountered when deprotection of compound 51, prepared from 39 by PDC oxidation, was attempted (Scheme 15).

Scheme 15

The starting point of our work during the third and final year of this contract was the synthesis of 2-formylinosine 53. The immediate precursor for the synthesis of this rare nucleoside was 2-vinylinosine 26 prepared as previously described from the reaction of the 2-iodo compound 19 with vinyltributyltin under palladium catalysis, followed by deprotection with trimethylsilyl iodide (Scheme 16). When 2-vinylinosine was subjected to ozonolysis under carefully controlled conditions and the reaction mixture reductively worked up, 2-formylinosine (53), was isolated in about 50% yield. This target compound was purified by reversed-phase HPLC on Amberlite XAD-4 resin with water-ethanol as the eluting solvent. The overall yield of 53 from guanosine was 14%. This compound was fully characterized by UV, FTIR, high-field $^{1}\mathrm{H}$ and $^{13}\mathrm{C}$ NMR, and high resolution fast atom bombardment mass spectrometry (FAB HRMS). The compound exists in both the formyl and its hydrated form and evidence for the presence of both forms can be easily discerned from the carbon spectrum. The chemical shift of the aldehyde carbon at about 185 ppm is consistent with the amide-like character of this carbonyl group.

H₂C=CH N
$$\frac{O_3}{MeOH,-65°C}$$
 $\frac{O_3}{MeOH,-65°C}$ $\frac{H_2O}{HOOH}$ $\frac{H_2O}{HOOH}$ $\frac{H_2O}{HOOH}$ $\frac{H_2O}{HOOH}$ $\frac{H_3O}{HOOH}$ $\frac{H_3O}{HOOH}$ $\frac{1}{1000}$ $\frac{1}{10000}$ $\frac{1}{1000}$ $\frac{1}{10000}$ $\frac{1}{10000}$ $\frac{1}{1000}$ $\frac{1}{10$

The key precursor for the synthesis of 2-hydroxymethylinosine (56) was the 2-vinyl compound 38 (Scheme 17). Controlled ozonolysis of compound 38 followed by reductive work-up with dimethyl sulfide gave the 2-formylinosine 54 in 54% yield. Reduction of 54 with sodium borohydride gave the alcohol 55 in 96% yield. Deprotection of 55 with trimethylsilyl iodide followed by tetraethylammonium fluoride gave the target compound 56. Target molecule 56 was purified by reversed-phase HPLC on Amberlite XAD-4 resin with water-ethanol as the eluting solvent. The overall yield of 56 starting from guanosine was about 18%. This compound was characterized by UV, FTIR, high-field H and 13C NMR, and high-resolution fast atom bombardment mass spectrometry (FAB HRMS).

$$CH_{2}CI_{2}$$

$$-67 °C$$

$$NaBH_{4}$$

$$TMSiCI$$

$$CH_{3}CN$$

$$R$$

$$NaBH_{4}$$

$$TMSiCI$$

$$CH_{3}CN$$

$$Et_{4}NF$$

$$CH_{3}CN$$

$$R$$

$$Et_{4}NF$$

$$CH_{3}CN$$

$$R$$

$$S_{6}$$

$$CH_{3}CN$$

$$R$$

$$S_{7}$$

$$S_{8}$$

$$S_{8}$$

$$S_{1}$$

$$S_{1}$$

$$S_{2}$$

$$S_{3}$$

$$S_{4}$$

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$$S_{1}$$

$$S_{1}$$

$$S_{2}$$

$$S_{3}$$

$$S_{4}$$

$$S_{5}$$

$$S_{6}$$

$$S_{7}$$

$$S_{8}$$

Scheme 17

2-Vinylnebularine (27) is the immediate precursor for the synthesis of the novel nebularine analogue, 2-formylnebularine (57). Ozonolysis of 27 under carefully controlled conditions followed by reductive work up gave 2-formylnebularine (57) in 61% yield (Scheme 18). This target molecule was purified by reversed-phase HPLC and fully characterized. The overall yield of 57 starting from guanosine was 11%.

The immediate precursor for the synthesis of the C-2 functionalized alcohol 59, also submitted for antiviral evaluation during the third year, is the 2-keto purine ribonucleoside 43, prepared from silylated 2-iodo-6-methoxypurine ribonucleoside 20 by a palladium-catalyzed cross-coupling reaction with tributyltin methoxide and 2-pentene-3-acetate. Reduction of 43 with sodium borohydride gave the diastereoisomeric alcohols 58 in 77% yield. Deprotection of 58 with trimethylsilyl iodide followed by tetraethylammonium fluoride and purification of the resulting product by HPLC gave the target molecule 59 (Scheme 19, 11% overall yield from guanosine).

Scheme 19

In the third year, we were also successful in synthesizing the 2-carbox-amidonebularine 64. This compound was synthesized by a methodology different from that used for the previous two compounds. A key starting compound for this synthesis was 8. When the 2-amino compound 8 was treated with n-pentyl nitrite and dimethyl disulfide in acetonitrile, thioalkylation occurred to give 60 as the product in 41% yield (Scheme 20). Oxidation of 60 with oxone proceeded smoothly to give the sulfoxide 61 in 75% yield. The S_{RN}l reaction of 61 with cyanide ion followed by deprotection gave 2-cyanonebularine (63) in 30% overall yield. 2-Cyanonebularine is a new nucleoside. Hydrolysis of this compound (NaOH, H₂O₂, EtOH) gave the carboxamide 64 in 31% yield. The overall yield of this new nucleoside starting from guanosine was 1%. It was purified, characterized, and submitted for antiviral evaluation.

Scheme 20

Synthesis of the novel carboxamide 70 was carried out starting with compound 60. Deprotection of 60 with sodium methoxide in methanol and subsequent oxidation with oxone converted 60 in to the sulfone 66 (25% overall yield from 8). Silylation of 66 (72%) followed by an S_{RN}1 reaction with the anion of diethyl malonate resulted in the formation of 68 (72%) (Scheme 21). It appears that 67 is initially converted to the diethyl malonyl derivative which undergoes a base-catalyzed retro-Claisen to 68. Deprotection of 68 (81%) followed by reaction of the resulting compound (69) with methanolic ammonia gave the target amide 70 in 75% yield after purification by reversed-phase HPLC. Compound 70 was characterized by spectroscopic methods and submitted for antiviral evaluation.

Synthesis of the target molecule, 2-acetamidoinosine 77, was first attempted via the base-catalyzed hydrolysis of the precursor nitrile, synthesized from the interesting palladium-catalyzed cross-coupling reaction between 20 and tri-n-butyl(cyanomethyl)stannane. However, this hydrolysis was unsuccessful under a variety of conditions. An alternative approach, however, was completely successful (Scheme 22). The starting compound for this approach was the 2-amino nucleoside 7. A thermal radical deamination thioalkylation of 7 followed by deprotection with sodium methoxide in methanol and subsequent oxidation with oxone converted 7 in about 38% overall yield to the sulfone 73. Silylation of 73 (63%) followed by an S_{RN} 1 reaction with the anion of diethyl malonate resulted in the formation of the 2-malonate of 74. Reaction of the latter with sodium ethoxide in refluxing ethanol gave 75 in 60% yield (for the two steps). Deprotection of 75 (42%) followed by reaction of the resulting compound (76) with methanolic ammonia gave the target amide 77 in 50% yield after purification by reversed-phase HPLC. Compound 77 was characterized by UV, FTIR, FAB HRMS, and NMR data and submitted.

Scheme 21

The key precursor for the synthesis of E-2-(1-propenyl)inosine 80 was the 2-iodo-6-methoxypurine nucleoside 19 (Scheme 23). Treatment of 19 with bis(acetonitrile)palladium II chloride with allyl tri-n-butylstannane in DMF at 100 °C for 6h, afforded a mixture of the 2-ally1-6-methoxypurine [2-(2-propenyl)-6-methoxypurine] 78, and the rearranged product, 2-(1propenyl)-6-methoxypurine 79 in a ratio of 2.5 to 1 and a combined yield of 92%. It is likely that the isomerization of the 2-propenyl to the 1-propenyl group is occurring at the slow step of the reaction, i.e. the transmetalation Temperature appears to be an important factor in these reactions. Below 90 °C, the reaction is extremely sluggish. The formation of the rearranged product is minimum at 90 °C. On the other hand, exclusive formation of the 1-propenyl compound 79 is realized when the temperature is raised to 110 °C. Deprotection of 79 with trimethylsilyl iodide in acetonitrile/DMF gave the target compound 80 in 81% yield after purification by reversed-phase HPLC on Amberlite XAD-4 resin. Complete structural characterization was carried out by UV, FTIR, FAB HRMS, and NMR data. high-field H NMR spectrum (in DMSO-d₆) gave unequivocal evidence for the Estereochemistry of the exocyclic double bond (J=17.0 Hz).

Scheme 22

Scheme 23

6. List of Target Compounds and Intermediates Submitted:

(i) 2-Acetonylinosine or 2-Acetonyl-9-(\beta-D-ribofuranosyl)hypoxanthine

AVS Identifying Number: AVS-002159

Contractor's Identifying Code No: VN-I-101

Final Report Reference: Scheme 5

(ii). 2-(2-Hydroxypropyl)inosine or 2-(2-Hydroxypropyl)-9-(3-D-ribofuranosyl)hypoxanthine

AVS Identifying Number: AVS-002352

Contractor's Identifying Code No: VN-I-102

Final Report Reference: Scheme 5

(iii) 2-Vinyl-9-(β -D-ribofuranosyl)hypoxanthine or 2-Vinylinosine

AVS Identifying No: AVS-002716

Contractor's Identifying Code No: VN-I-103

(iv) 2-Vinyl-9-(β -D-ribofuranosyl)purine or 2-Vinylnebularine

AVS Identifying No: AVS-002694

Contractor's Identifying Code No: VN-I-104

Final Report Reference: Scheme 8

(v) $2-(1,2-Dihydroxyethy1)-9-(\beta-D-ribofuranosy1)$ hypoxanthine or 2-(1,2-Dihydroxyethy1)inosine

AVS Identifying No: AVS-002695

Contractor's Identifying Code No: VN-I-105

Final Report Reference: Scheme 9

(vi) 2-(1,2-Dihydroxyethyl)-9-(\beta -D-ribofuranosyl)purine or 2-(1,2-Dihydroxyethyl)nebularine

AVS Identifying No: AVS-002883

Contractor's Identifying Code No: VN-I-106

(vii) 2-(2-Hydroxyethyl)-9-(β -D-ribofuranosyl)hypoxanthine or 2-(2-Hydroxyethyl)inosine

AVS Identifying No: AVS-002884

Contractor's Identifying Code No: VN-I-107

Final Report Reference: Scheme 11

(viii) 2-Acetonyl-9-(β -D-ribofuranosyl)purine or 2-Acetonylnebularine

AVS Identifying No: AVS-003039

Contractor's Identifying Code No: VN-I-108

Final Report Reference: Scheme 6

(ix) 2-(2-Hydroxypropyl)-9-(β -D-ribofuranosyl)purine or 2-(2-Hydroxypropyl)nebularine

AVS Identifying No: AVS-003582

Contractor's Identifying Code No: VN-I-109

(x) $2-Iodo-9-(\beta-D-ribofuranosyl)$ purine or 2-Iodonebularine

AVS Identifying No: AVS-003923

Contractor's Identifying Code No: VN-I-110

Final Report Reference: Page 11

(xii) 2-(1-Methyl-2-oxobutyl)-9-(β -D-ribofuranosyl)purine

AVS Identifying No: AVS-003924

Contractor's Identifying Code No: VN-I-111 Final Report Reference: Scheme 13

(xii) 2-(1-Methyl-2-oxobutyl)-9-(\$\beta\$-D-ribofuranosyl)hypoxanthine

AVS Identifying No: AVS-003921

Contractor's Identifying Code No: VN-I-112

(xiii) 2-Iodo-9-(\$\beta\$-D-ribofuranosyl)hypoxanthine or 2-Iodoinosine

AVS Identifying No: AVS-003922

Contractor's Identifying Code No: VN-I-113

Final Report Reference: Page 11

(xiv) 2-Formylinosine

AVS Identifying No: AVS-004094

Contractor's Identifying Code No: VN-I-114

Final Report Reference: Scheme 16

(xv) 2-(E-2-Ethoxyvinyl)nebularine

AVS Identifying No: AVS-004095

Contractor's Identifying No: VN-I-115

(xvi) 2-(2-Hydroxy-1-methylbuty1)-9-(/3 -D-ribofuranosyl)hypoxanthine

AVS Identifying No: AVS-004109

Contractor's Identifying Code No: VN-I-116

Final Report Reference: Scheme 19

(xvii) 2-Hydroxymethylinosine

AVS Identifying No: AVS-004232

Contractor's Identifying Code No: VN-I-117

Final Report Reference: Scheme 17

(xviii) 2-Formylnebularine

AVS Identifying No: AVS-004233

Contractor's Identifying Code No: VN-I-118

(xix) 2-Carboxamidonebularine

AVS Identifying No: AVS-004528

Contractor's Identifying Code No: VN-I-119

Final Report Reference: Scheme 20

(xx) $E-2-(1-Propeny1)-9-(\beta-D-ribofuranosyl)$ hypoxanthine

AVS Identifying No: AVS-004727

Contractor's Identifying Code No: VN-I-120

Final Report Reference: Scheme 23

(xxi) 2-Acetamidonebularine

AVS Identifying No: AVS-004928

Contractor's Identifying Code No: VN-I-121

(xxii) 2-Acetamidoinosine

AVS Identifying No: Contractor's Identifying Code No: VN-I-122 Final Report Reference:

7. Antiviral Screening Data:

AVS Identifying Number Contractor's Code Number Antiviral Drug Screening
Data

AVS-002159 VN-I-101

Very active, specific
TI >1000 (in vitro, SF)
HIV results not available
Toxic in CCHF suckling mouse

AVS-002352 VN-I-102

Some activity against YF
Not active against VSV, AD2, VV,
HIV, SF, JE <u>in vitro</u>
<u>in vivo</u> data not available

AVS-002716 VN-I-103 Some broad spectrum activity (in vitro) against YF, JE, AD2, VV, PT, RVF, not active against VEE, VSV, HIV
In vivo data not available

AVS-002694 VN-I-104 Not active (<u>in vitro</u>)
<u>In vivo</u> data not available

AVS-002695 VN-I-105 Not active (<u>in vitro</u>)
<u>In vivo</u> data not available

AVS-002883 VN-I-106

Some activity against RVF
Not active (<u>in vitro</u>)
against other viruses
HIV results not available
<u>In vivo</u> data not available

AVS-002884	Some activity against RVF
VN-I-107	Not active (<u>in vitro</u>)
	against other viruses
	HIV results not available
	<u>In vivo</u> data not available
AVS-003039	Not active against AD2, JE,
VN-I-108	VSV, VV, RVF (in vitro)
V.N. 2 100	In vivo data not available
	in vivo data not available
AVS-003582	Not active against JBE, VV, AD2,
VN-I-109	VSV, RVF (in vitro)
VII 2 10)	In vivo data not available
	in vivo data not available
AVS-003923	Not active against VSV, AD2, VV,
VN-I-110	SFS, VEE, JBE, YF, RVF (in vitro)
VN-1-110	
	<u>In vivo</u> data not available
AVS-003924	Not active against VSV, VV, AD2,
VN-I-111	RVF (in vitro)
VN-1-111	
	<u>In vivo</u> data not available
AVS-003921	Not active against VSV, AD2, VV,
VN-I-112	SFS, VEE, JBE, YF, RVF (in vitro)
VN-1-112	
	<u>In vivo</u> data not available
AVS-003922	Not active against VSV, VV, AD2,
VN-I-113	
AN-1-112	JBE, VEE, SFS, YF, RVF (in vitro)
	<u>In vivo</u> data not available
AVS-004094	Some activity against AD2, VSV
VN-I-114	Not active against JBE, PT, SFS,
410-1-114	
	VEE, VV, YF (in vitro)
	<u>In vivo</u> data not available
AVS-004095	Not active against VSV, AD2, VV,
VN-I-115	YF, VEE, PT, SFS, JBE (in vitro)
VN-1-115	Tr, VEE, FI, SFS, SBE (III VICTO)
	<u>In vivo</u> data not available
AVS-004109	Not active against VSV, AD2, VV,
VN-I-116	YF, VEE, PT, SFS, JBE (in vitro)
VN-1-110	
	<u>In vivo</u> data not available
AVS-004232	Some activity against VV (in vitro)
VN-I-117	Not active against AD2, JBE, VSV
1.1 - 11/	
	<u>In vivo</u> data not available
AVS-004233	Not active against AD2, JBE, VSV,
VN-I-118	VV (in vitro)
1M T 110	
	<u>In vivo</u> data not available
AVS-004528	Not active against AD2, VV, VSV
VN-I-119	(in vitro)
1M T-117	
	<u>In vivo</u> data not available

AVS-004727 VN-I-120 Not active against AD2, VV (<u>in vitro</u>)
Other data not available

AVS-004928 VN-I-121 Screening data not available

AVS-VN-I-122 Screening data not available

8. Bibliography of Publications, Patents, and Presentations:

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- (iii) V. Nair, S. D. Chamberlain, R. DeSilvia, Jr., and G. S. Buenger, Synthetic Approaches to Rare 2-Substituted Purine Nucleosides, <u>Nucleosides and Nucleotides</u>, 1987, 6, 229 (4 copies furnished to SGRD-RMS).
- (iv) V. Nair and D. A. Young, Conformational Correlation of Purine Nucleosides by High-Field Carbon-13 NMR Data, <u>Magnetic Resonance in Chemistry</u>, 1987, 25, 937 (4 copies furnished to SGRD-RMS).
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- (x) V. Nair and G. S. Buenger, Novel 2-Substituted Purine Nucleosides, 99th Session of the Iowa Academy of Science, Grinnell, Iowa, April, 1987.
- (xi) V. Nair and A. G. Lyons, Functionalization of Inosine, 99th Session of the Iowa Academy of Science, Grinnell, Iowa, April, 1987.
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- (xiv) V. Nair, <u>Planery Lecture</u>, 8th International Symposium on Nucleosides, Nucleotides, and their Biological Applications, Perdido Beach, Alabama, October, 1988.
- (xv) V. Nair and A. G. Lyons, Synthesis of Novel Purine Nucleoside Carboxaldehydes, 23rd Midwest Regional Meeting of the American Chemical Society, Iowa City, November, 1988.
- (xvi) V. Nair, G. A. Turner, G. S. Buenger and S. D. Chamberlain, New Methodologies for the Synthesis of C-2 Functionalized Hypoxanthine Nucleosides, <u>Journal of Organic Chemistry</u>, 1988, <u>53</u>, 3051 (4 copies furnished to SGRD-RMS)
- (xvii) V. Nair and G. S. Buenger, Rare Purine Nucleosides: Congeners of the Antibiotic, Nebularine, <u>Synthesis</u>, 1988, 848 (4 copies furnished to SGRD-RMS)
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9. Personnel Supported:

Stanley D. Chamberlain, Ph.D. Degree, December 1986.
Raymond DeSilvia, Jr., M.S. Degree, August 1986 (Deceased).
Gregory A. Turner, Ph.D. Degree, May, 1987
Greg S. Buenger, Ph.D. Degree (Expected May 1989)
Arthur G. Lyons, Ph.D. Degree (Expected May 1989)
Brian J. Hettrick, M. S. Degree, May, 1988

10. Summary:

In the over three years on this contract, we have had considerable success in our synthetic work and a total of twenty-two rare 2-substituted purine nucleosides were synthesized, purified, characterized, and submitted to the Department of Antiviral Studies with supporting data. Although screening data are not complete as yet on several of the compounds submitted, some very One compound (2interesting and positive data have been received. acetonylinosine, AVS-00159) has been found to have very high activity (TI > 1000) against the Sandfly Fever Virus (Phlebovirus). Another compound (2vinylinosine, AVS-002716) has been found to have low but broad spectrum activity against a number of RNA viruses. Two other compounds (AVS-002883 and AVS-002884) have shown some activity against the Rift Valley Fever Virus, and still another (AVS-002352) has shown activity against the Yellow Fever Virus. 2-Formyinosine (AVS-004094) has shown some activity against Type 2 Adenovirus and 2-hydroxymethylinosine (AVS-004232) has shown activity against the Vaccinia Virus. Ten publications and a patent have arisen directly from this Various aspects of the synthetic work have also been presented as invited and contributed papers (including a planery lecture) and research seminars at regional, national, and international scientific meetings and occasions.